**Introduction**

Intraspinal nerve block is a kind of anesthesia method that is often used clinically, with the advantages of safety and less equipment. It is also characterized with simple operation, small medical expense, better effect of analgesia and muscle relaxation, controllable characteristics as well as the maintenance of the patient’s consciousness (1). However, it also has shortcomings, such as hypotension, tachycardia, headache after puncture, damage to blood vessels and nerves, spinal cord injury (SCI) and so on (2). Although the incidence of serious complications is relatively low, it is more difficult to recover neurological function in the event of severe neurological complications (3). Recently, due to the reports on the occurrence of neurological complications caused by the spinal canal block after local anesthesia, intraspinal block has gained more attention (4).

A large number of studies have shown that SCI can cause the release of local inflammatory cytokine, which may be associated with neurological dysfunction (5). In order to reduce the direct SCI caused by puncture, many scholars have concentrated on the research and development on the lumbar puncture needle for many years, making improvement from the tip shape and needle diameter size...
respectively (6). The growth cone is the end of the neuronal axon, which plays an important role in the growth of nerve cells (5). Therefore, the study of the growth cone can provide good observation of the effects of some external stimuli or external substances on the development of nerve tissue (7).

For more than 30 years, Circular RNA served a crucial role in the occurrence and development of human diseases and had been acknowledged as products of aberrant splicing, gene rearrangement, or non-linear reverse splicing (8). MicroRNAs are 19–24 nucleotides in length, small noncoding RNAs, which posttranscriptionally regulate gene expression by targeting the 3’untranslated region (3’-UTR) of target mRNAs (9,10). With the profound research on miRNA, these small molecules are found to play important roles in the pathophysiology of the cardiovascular system, including arrhythmias, cardiac hypertrophy, fibrotic heart failure and angiogenesis (11). Therefore, the aim of this study was to analyze the effects of Circ 0000962 attenuates toxicity in SCI and its possible mechanism.

**Methods**

**Animals**

Forty Sprague-Dawley rats (6 weeks old, male) supplied by Animal Experimental Center of Medicine, University were randomly distributed into two groups: the sham group, SCI model group. Briefly, in SCI model group, the rats were fixed in the prone position and anesthetized by 35 mg/kg of pentobarbital. Skin was cut to 2–3 cm middle incision in the back, and vertebral T7–T9 were exposed. After stabilizing vertebral T7 and T9, a laminectomy was performed at the thoracic level T8. A syringe needle was used to induce the injury, which was released from a height of 12.5 mm above the surface of the cord, inflicting a moderate contusion. Hemostatic suture was performed layer by layer, and alcohol was then applied for disinfection. Three days after the operation, the rats received an intramuscular injection of 1 g gentamicin to prevent infection. The stitches were taken out after one week (Figure 1). Rats in the sham group were performed with laminectomy of vertebral T8 without injury. This study was approved by the Animal Experiment Committee of the First Affiliated Hospital of USTC, Division of Life Sciences and Medicine, University of Science and Technology of China.

**Basso, Beattie, and Bresnahan (BBB) locomotor rating score and water content of SCI**

BBB locomotor rating score was performed after treatment with Circ 0000962. BBB locomotor rating score was scored in an open field according to the locomotor rating scale of 0 (complete paralysis) to 21 (normal locomotion). After treatment with Circ 0000962, rat was sacrificed under 35 mg/kg of pentobarbital and Spinal cord tissue was weighed as wet weighing. Spinal cord tissue was dried at 70 °C for 72 h and weighed as drying weighing. Water content of SCI was calculated using wet weighing (g)/drying weighing (g) × 100%.

**Cell culture**

Nerve cell lines AGE1.HN cell were cultured in DMEM medium (Gibco, USA) supplemented with 10% fetal bovine serum (FBS, Gibco) at 37 °C in a humidified atmosphere with 5% CO₂. Circ 0000962, anti-Circ 0000962 and negative mimics were transfected into cell
using Lipofectamine 2000 (Thermo Fisher Scientific, Inc. Waltham, MA, USA). After transfection at 48 h, cell was treated with LPS (100 ng/mL) for 4 h.

**Measurement of inflammation and caspase-8/9 activity**

Serum of rats or cell samples were collected after 2,000 g for 10 min at 4 °C, and was used to measure TNF-α, IL-1β, IL-6 and IL-18 levels using ELISA KITS. Cell was treated for 6 h, supernatants of cell were collected after 2,000 g for 10 min at 4 °C, and was used to measure TNF-α, IL-1β, IL-6 and IL-18 levels using ELISA KITS. Absorbance was measured using ELISA reader at 450 nm.

**Western blot analysis**

Tissue samples or cell samples were homogenated or lysed using RIPA assay and protein content was measured using BCA assay. And 50 μg proteins was then subjected to 8–12% SDS-PAGE and transferred onto a polyvinylidene fluoride membrane (Merck KGaA, Darmstadt, Germany). The membrane was blocked in 5% non-fat milk in Tris-buffered saline/Tween 20 (TBST) for 1 h, and subsequently incubated with NF-κB, PI3K, p-Akt and GAPDH (Cell Signaling Technologies, Danvers, MA, USA) at 4 °C overnight. The membrane was washed with TBST and then incubated with horseradish peroxidase labeled goat anti-rat secondary antibody (1:1,000; Cell Signaling Technologies, Danvers, MA, USA) at 37 °C for 1 h. Protein blank was detected with enhanced chemiluminescence (ECL) assay.

**Statistical analysis**

Data are expressed as mean ± SD. Student’s t-test or ANOVA at P<0.05 was considered statistically significant. P<0.05 was considered to indicate a statistically significant difference.

**Results**

**Circ 0000962 in inflammation in SCI model**

Circ 0000962 was firstly investigated in our lab, therefore, we explored the function and mechanism of miRNA-203a in SCI model. In model of rats, TNF-α, IL-1β, IL-6 and IL-18 levels were increased in SCI modal rat, compared with sham group (Figure 2A,B,C,D). HE staining showed that the number of Spinal cord cells in SCI model group was lower than that of sham group (Figure 2E). We found that Circ 0000962 expression was decreased in SCI model rat, compared with sham group (Figure 2F,G).

**Circ 0000962 regulates inflammation in vitro model of SCI**

To analyze the function of Circ 0000962 on inflammation in SCI model, Circ 0000962 or anti-Circ 0000962 mimics regulates Circ 0000962 expression in vitro model. As shown in Figure 3A, Circ 0000962 inhibitor decreased the expression of Circ 0000962 in vitro model, compared with negative mimics group. Down-regulation of Circ 0000962 increased TNF-α, IL-1β, IL-6 and IL-18 levels in vitro model, compared with negative group (Figure 3B,C,D,E). Then, 0000962 mimics increased the expression of Circ 0000962 expression in vitro model, compared with negative group (Figure 3F). Over-expression of Circ 0000962 reduced TNF-α, IL-1β, IL-6 and IL-18 levels in vitro model, compared with negative group (Figure 3G,H,I,J).

**Circ 0000962 regulates inflammation in vitro model of SCI by miR-302b-3p**

To explain the mechanism of Circ 0000962 on inflammation in vitro model of SCI by miR-302b-3p. MiR-302b-3p expression was increased in SCI model, compared with sham group (Figure 4A,B,C). Then, over-expression of Circ 0000962 promoted miR-302b-3p expression in vitro model of SCI, compared with negative group (Figure 4D). 3’UTR region of Circ 0000962 showed potential alignment with miR-302b-3p sequence and luciferase reporter activity levels were reduced by over-expression of Circ 0000962, compared with negative group. There was increase the expression of miR-302b-3p in over-expression of miR-302b-3p, compared with negative group (Figure 4E). Over-expression of miR-302b-3p increased TNF-α, IL-1β, IL-6 and IL-18 levels, compared with negative group (Figure 4F,G,H,I). Then, anti-miR-302b-3p mimics reduced the expression of miR-302b-3p, and inhibited TNF-α, IL-1β, IL-6 and IL-18 levels, compared with negative group (Figure 4J,K,L,M,N).

**MiR-302b-3p regulates inflammation in vitro model of SCI via PI3K/Akt/NF-κB signaling**

To confirm the mechanism of miR-302b-3p on
inflammation in vitro model of SCI, the signaling was measured using gene chip. Gene chip showed that Overexpression of miR-302b-3p increased p65 and reduced PI3K expression in vitro model, compared with negative group (Figure 5A). Figure 5B,C showed that 3'UTR region of PIK3CA showed potential alignment with miR-302b-3p sequence and luciferase reporter activity levels were reduced by over-expression of miR-302b-3p, compared with negative group. Over-expression of miR-302b-3p suppressed PIK3CA expression in vitro model, compared with negative group (Figure 5D). As shown in Figure 5E,F,G,H, PI3K and p-Akt protein expressions were reduced, and NF-κB expression was increased in vitro model by over-expression of Circ 0000962, compared with negative mimics group. Over-expression of Circ 0000962 induced PI3K and p-Akt protein expressions and suppressed NF-κB protein expressions in vitro model, compared with negative mimics group (Figure 5I,J,K,L).

**Discussion**

In recent years, various treatments have focused on axonal
and dendritic repair to enhance recovery from CNS injury. SCI is medically and socioeconomically enervating and lacking in effective therapies. Improvements in the neurosciences have drawn more attention to SCI research, and until now, no available treatments for SCI could effectively decrease or stop the disease progression. The elucidation of the pathophysiological mechanisms underlying SCI is necessary for studying effective therapies for SCI. The development of SCI pathology is guided by the two leading mechanisms of damage, which are primary injury and secondary injury (4). Primary injury occurs passively within a short period of time after injury, resulting in irreversible damage, while secondary injury is reversible, controllable and complicated with self-destructive cascade reaction process, which is the main reason for aggravating neurological dysfunction (12). The research on miRNAs may provide new insights for the study of the molecular mechanisms of SCI. In this study, we identified that circ 0000962 expression was decreased in SCI model rat.

Figure 3 Circ 0000962 regulates inflammation in vitro model of SCI. (A) Circ 0000962 expression by down-regulation of Circ 0000962; (B) TNF-α levels by down-regulation of Circ 0000962; (C) IL-1β levels by down-regulation of Circ 0000962; (D) IL-6 levels by down-regulation of Circ 0000962; (E) IL-18 levels by down-regulation of Circ 0000962; (F) Circ 0000962 expression by over-expression of Circ 0000962; (G) TNF-α levels by over-expression of Circ 0000962; (H) IL-1β levels by over-expression of Circ 0000962; (I) IL-6 levels by over-expression of Circ 0000962; (J) IL-18 levels by over-expression of Circ 0000962. Negative, control negative group; Circ 0000962, over-expression of Circ 0000962 group; Circ 0000962 inhibitor, down-regulation of Circ 0000962 group. ##, P<0.01 compared with control group. Data are expressed as mean ± SD. Student's t-test at P<0.05 was considered statistically significant. SCI, spinal cord injury.
neurological deterioration of inflammatory cytokines could also cause neurons and oligodendrocyte apoptosis through a pathway to stimulate the proliferation of astrocytes, leading to local formation of glial paralysis, inhibition of axonal regeneration, upregulation of other inflammatory-related gene expression (6,13). A wide range of cytokines can be divided into inflammatory cytokines and anti-inflammatory cytokines according to the differences in inflammatory response (13). A variety of proinflammatory cytokines and chemokines are significantly increased in post-injury area, therefore, the post-inflammatory cascade is expanded to increase secondary damage to SCI (5). We found revealed that over-expression of Circ 0000962 decreased TNF-α, IL-1β, IL-6 and IL-18 levels in vitro model. miR-302b-3p expression was increased in SCI model. Wang et al. reported that knockdown miR-302b reduced LPS-induced injury by in C28/I2 chondrocytic cells (14), which was also in accordance with our results.

Many studies have indicated that the inhibition of NF-κB activation protects the spinal cord in SCI (15-17).
Figure 5 MiR-302b-3p regulates inflammation in vitro model of SCI via PI3K/Akt/NF-κB signaling. (A) Gene chip; (B) 3'UTR region of PIK3CA showed potential alignment with miR-302b-3p sequence; (C) luciferase reporter activity; (D) miR-302b-3p suppressed PIK3CA expression; (E) NF-κB protein expression by over-expression of Circ 0000962; (F) p-Akt protein expression by over-expression of Circ 0000962; (G) PI3K protein expression by over-expression of Circ 0000962; (H) western blotting assays by over-expression of Circ 0000962; (I) NF-κB protein expression by down-regulation of Circ 0000962; (J) p-Akt protein expression by down-regulation of Circ 0000962; (K) PI3K protein expression by down-regulation of Circ 0000962; (L) Western blotting assays by down-regulation of Circ 0000962. Negative, control negative group; Circ 0000962, over-expression of Circ 0000962 group; anti-203a, down-regulation of Circ 0000962 group. *P<0.01 compared with control group. Data are expressed as mean ± SD. Student’s t-test at P<0.05 was considered statistically significant.

Thus, NF-κB activation is the key element of the secondary neuronal damage in SCI. In the present study, PI3K and p-Akt protein expressions were increased, and NF-κB expression was reduced in vitro model by over-expression of Circ 0000962. NF-κB activity as well as TNF-a, IL-1b, and IL-6 expression was suppressed by Circ 0000962 treatment, indicating that NF-κB activation contributes to increased levels of these cytokines. Previous results and data showing the severity of neuronal injury were dependent on the expression of cytokines after a SCI (18).

PI3K/Akt signaling pathway plays an important role in neurophysiological pathology as a central controller for cell growth, proliferation, survival and differentiation under conditions such as extracellular signal, growth factor, cell energy status and emergency response (19,20). PI3K/Akt signaling pathway has been involved in the process of glial scar formation, and attenuation of PI3K/Akt pathway may beneficially inhibit glial scar formation after SCI. Guo et al. indicated the microRNA-302b-3p suppresses cell proliferation through AKT pathway in gastric cancer (21). Our results revealed that miR-302b-3p reduced the effects of circ 0000962 on inflammation in vitro model of SCI through PI3K/Akt/NF-κB-dependent signaling.

In conclusion, our study revealed that Circ 0000962 weakened inflammation of SCI through anti-inflammation effects by PI3K/Akt/NF-κB-dependent signaling in SCI by miR-302b-3p. This study contributes to our understanding of molecular mechanisms of Circ 0000962, and the investigation
Figure 6 MiR-302b-3p reduced the effects of Circ 0000962 on inflammation in vitro model of SCI through PI3K/Akt/ NF-κB signaling. (A) NF-κB protein expression; (B) p-Akt protein expression; (C) PI3K protein expression; (D) western blotting assays; (E) TNF-α levels; (F) IL-1β levels; (G) IL-6 levels; (H) IL-18 levels. Negative, control negative group; Circ 0000962, over-expression of Circ 0000962 group; PI3K, over-expression of Circ 0000962 group and PI3K group. **, P<0.01 compared with control group; ***, P<0.01 compared with Circ 0000962 group. Data are expressed as mean ± SD. ANOVA at P<0.05 was considered statistically significant.

on the new molecular therapeutic targets in SCI.

Acknowledgments
None.

Footnote
Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study was approved by the Animal Experiment Committee of the First Affiliated Hospital of USTC, Division of Life Sciences and Medicine, University of Science and Technology of China (No. 201711(16)).

References
5. Allison DJ, Josse AR, Gabriel DA, et al. Targeting


